

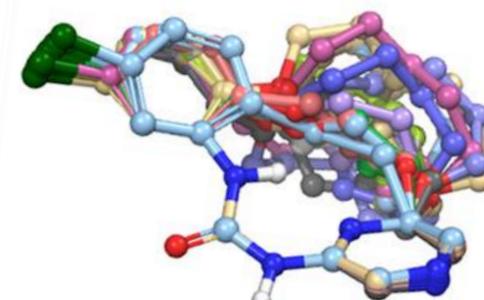
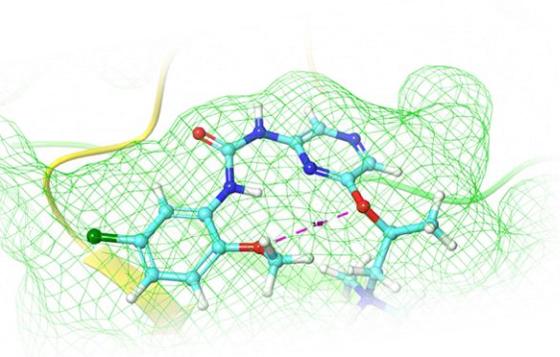
Conformational Sampling and Binding Affinity Prediction of Macrocycles



David Rinaldo, Dan Sindhikara

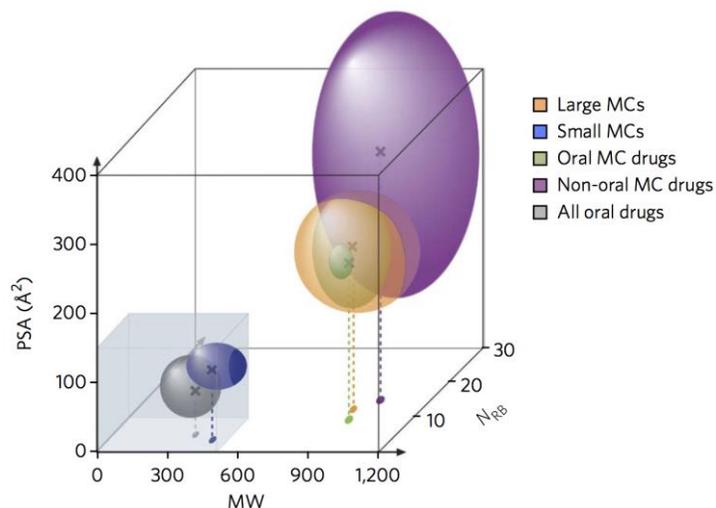
Strasbourg Summer School in Chemoinformatics

June 25th 2018



Macrocycles in Drug Design

- **Macrocycles are pervasive**
(63 on market, 35 in development)¹
- **Cyclization can impart conformational stability of known binder**
(linear compound, PPIs)
- **Cyclization can expand Ro5 restrictions²**



status	class	route of administration	N
<u>registered</u>	small molecule	oral	1589
		oral, MW > 500	89
	macrocycle	oral	18
		parenteral	45
	cyclic peptide	oral	1
		parenteral	26
macrolide	oral	14	
	parenteral	9	
<u>clinical development</u>	macrocycle	oral	15
		parenteral	20
	cyclic peptide	oral	3
		parenteral	8
	macrolide	oral	2
		parenteral	3
"de novo designed"	oral	9	
	parenteral	1	

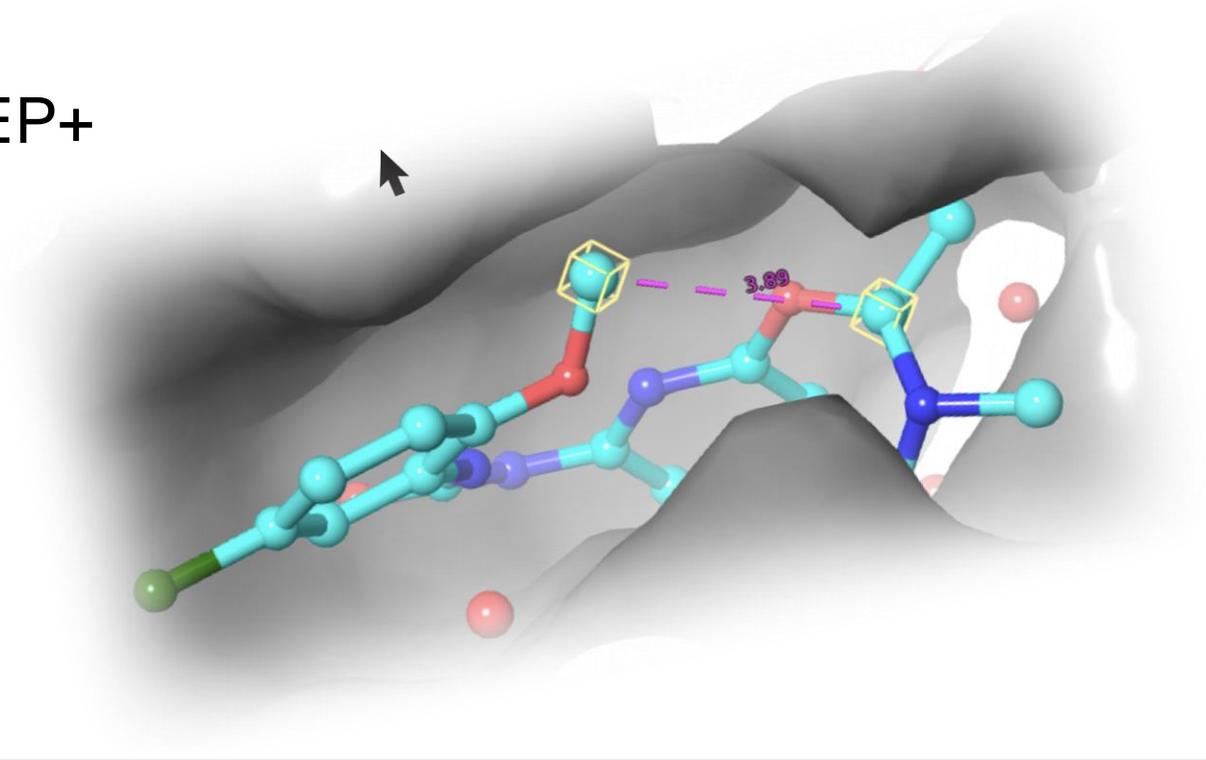
Challenges in Macrocycle Design

- **Difficult synthetically**
 - Cyclization reactions can be difficult, time-consuming
 - High MW can be burdensome
- **Difficult computationally**
 - Cyclization impedes sampling
 - Force field parameterization requires “unphysical” twisting of ring torsions
 - Chemical software not typically designed for large flexible rings
 - Design workflows must be adapted for macrocycle caveats

Schrodinger's Macrocyclic Design Solutions

Schrödinger is expanding the limits of *in silico* macrocycle design

- Prime macrocycle conformational sampling
- Bioactive conformer stability
- Docking with Glide
- Free Energy Perturbation with FEP+
- Passive membrane permeability



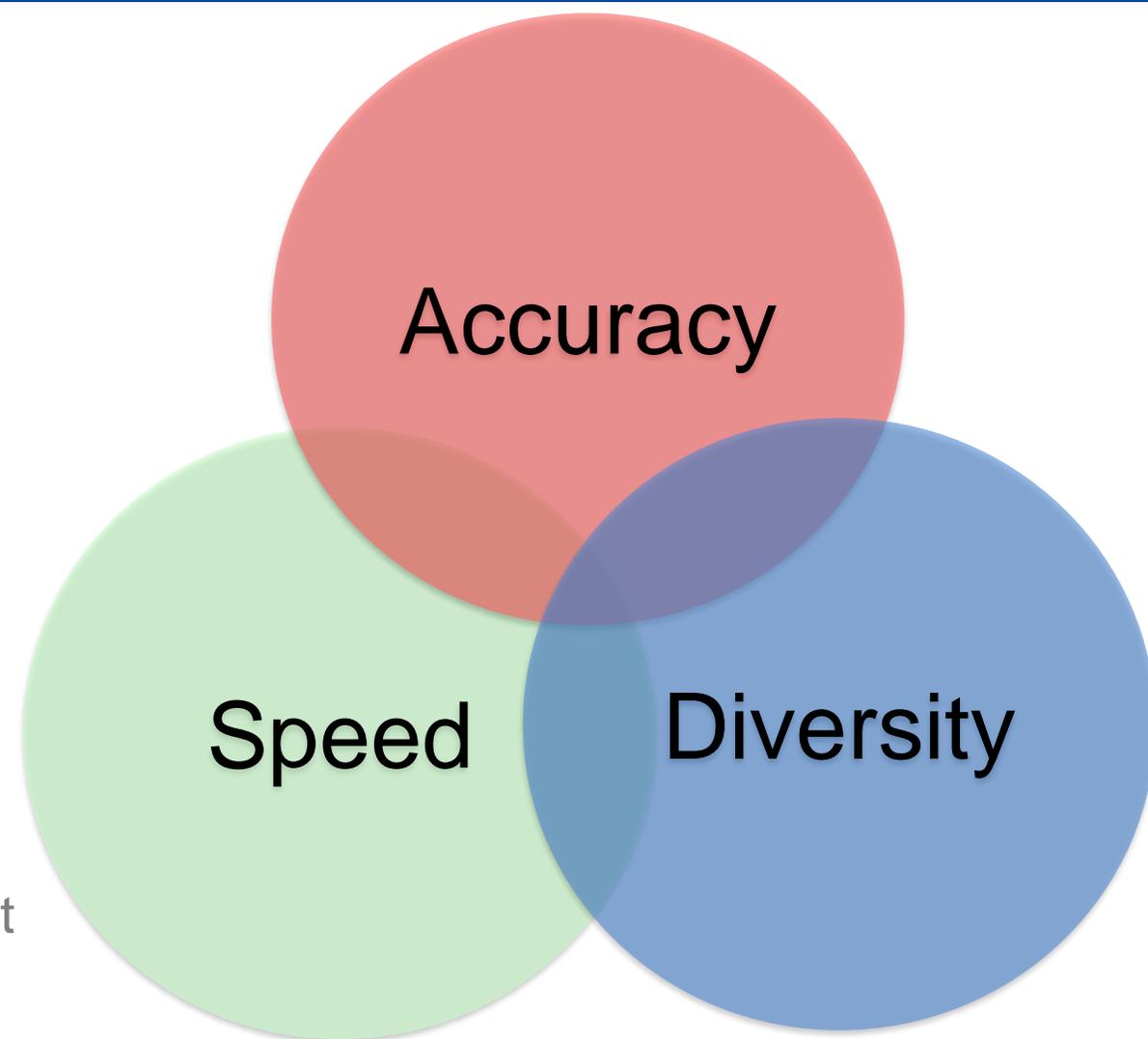
A molecular simulation visualization showing a protein backbone as a grey ribbon structure. A ligand molecule is bound within a pocket of the protein, represented by a cluster of orange spheres. The text "Macrocycle Sampling" is overlaid in white on the central part of the image.

Macrocycle Sampling

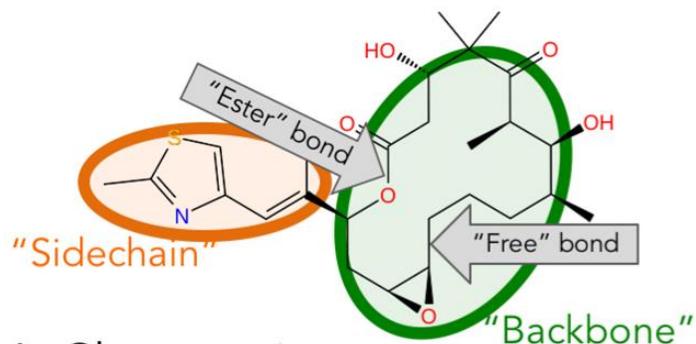
Sampling: Attributes of a functional sampler

Sampling algorithm designed to efficiently explore macrocycle conformational space, especially major ring conformations

- Accuracy
 - Match experimental conformations
- Speed
 - Fast enough to enable workflows
 - Quick turnaround during ideation
- Diversity
 - Sample not only crystal conformation, but also permeable, solvent, exploitable

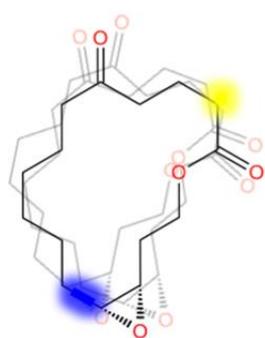


PrimeMCS Algorithm



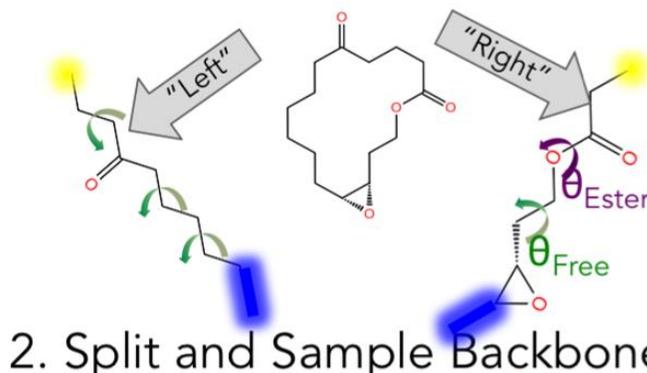
1. Characterize

Recognize macrocyclic topology, assign bond rotamers from library



3. Re-form and cluster backbone

Find pairs of half-loops which form reasonable closed rings. If there's enough, cluster them, otherwise, go back to 2 with finer resolution

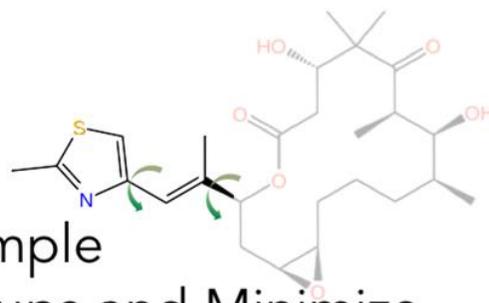


2. Split and Sample Backbone

Break* major ring, combinatorially sample "half-loops" using bond rotamers.

4. Sample R-groups and Minimize

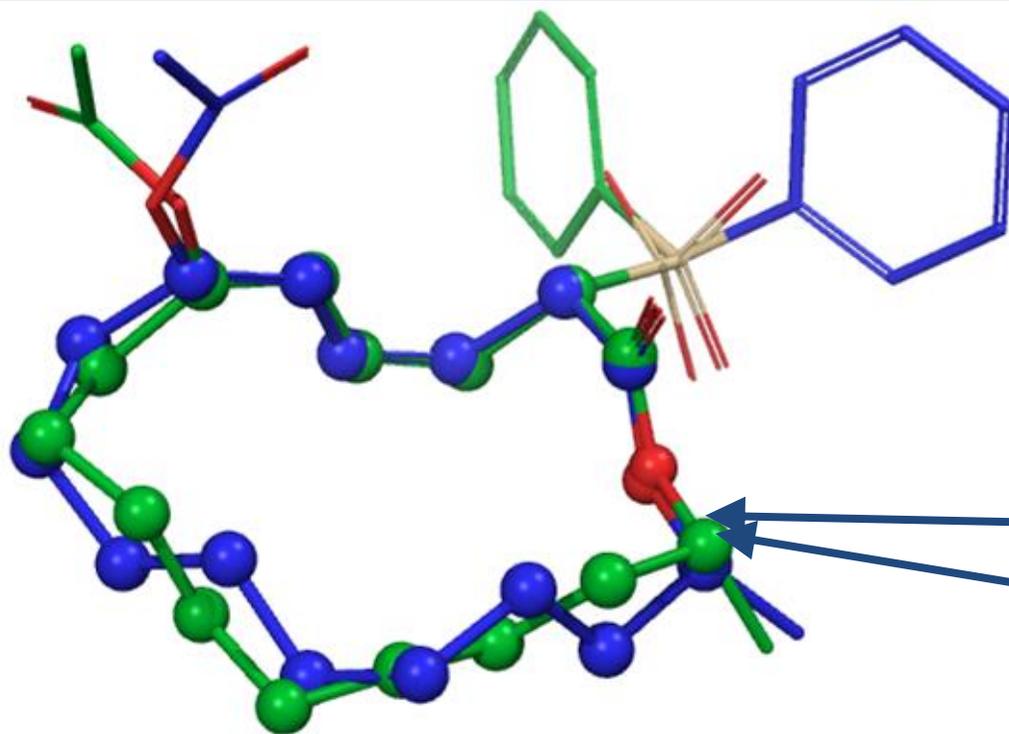
Build out r-groups from clustered ring backbones using bond rotamers. Minimize to relax backbone and repeat once.



Algorithm details of note

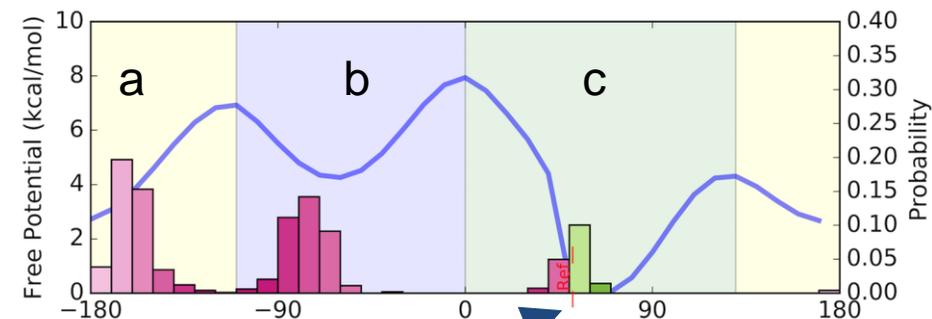
- Originally developed @ Prof Matt P. Jacobsons Lab based on loop sampling algorithm
- In step 2, we cut the ring 10 different ways around the ring (spinroot 10), enabling 10x parallelism
- Cross-links are accounted for via inverse clash restraints
- Only one independent ring system is thoroughly sampled

Measuring Macrocycle Backbone Structure



Backbone RMSD

Assuming that backbone sampling is the difficult part. Here we measure the best backbone RMSD of a structure in the ensemble to the crystal reference



cccbbbccaaca

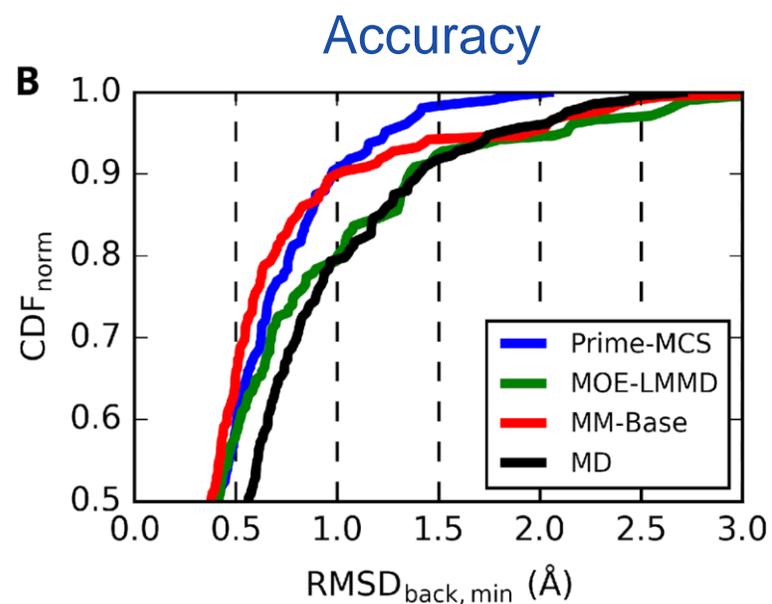
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Backbone Torsional Fingerprint

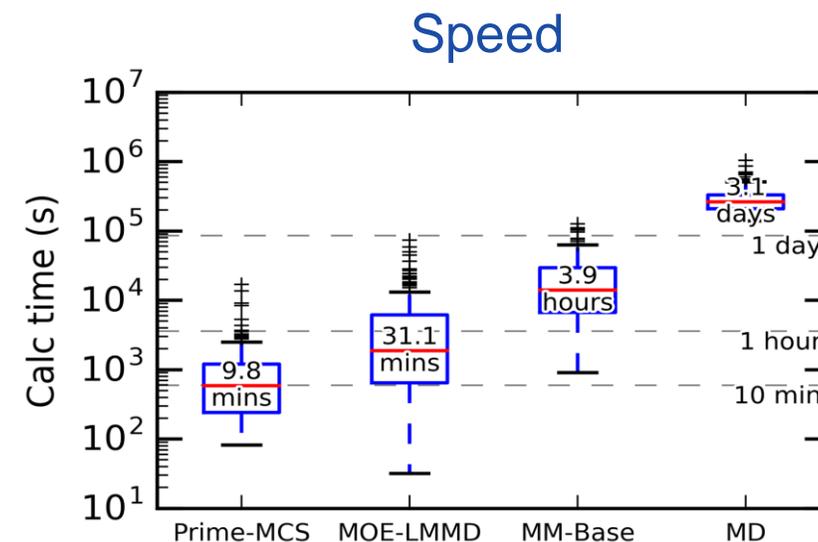
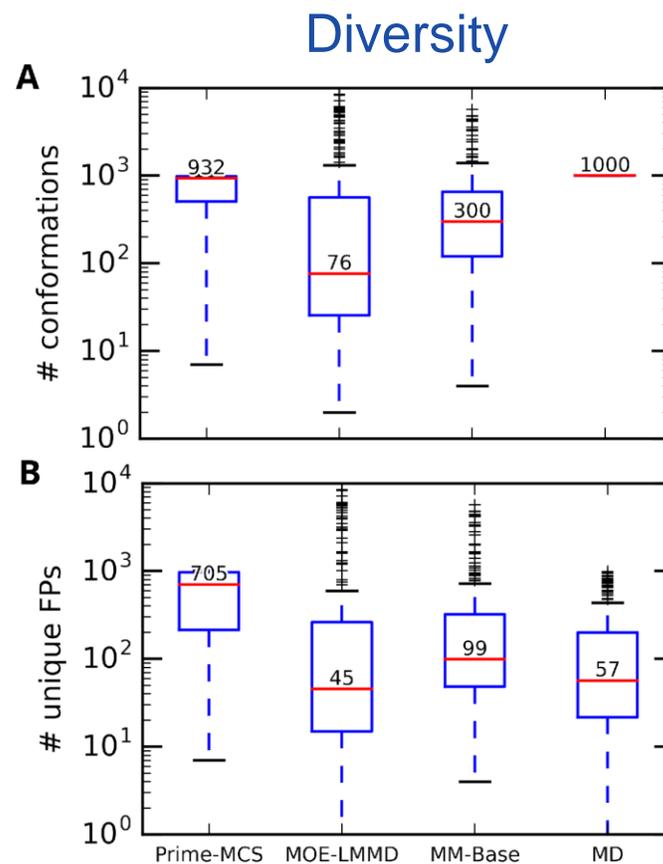
Using torsional scanning profiles, we can bin torsions into states. The superposition of all backbone states (torsional fingerprint) can be used to compare discretized conformations.

Measuring Accuracy, Diversity, and Speed

Test set consisting of 208 diverse crystal structures, Comparison of 4 different protocols ¹



PrimeMCS finds < 1.0Å structure
90% of the time, worst outlier 2.1Å



PrimeMCS jobs are 10x
parallelizable, enabling extremely
fast turnaround times.

A molecular simulation showing a protein backbone as a grey ribbon structure. A ligand is bound to the protein, represented by orange spheres and a blue and yellow stick model. The text "Predicting Stable Cyclizations of Linear Ligands" is overlaid in white on the protein structure.

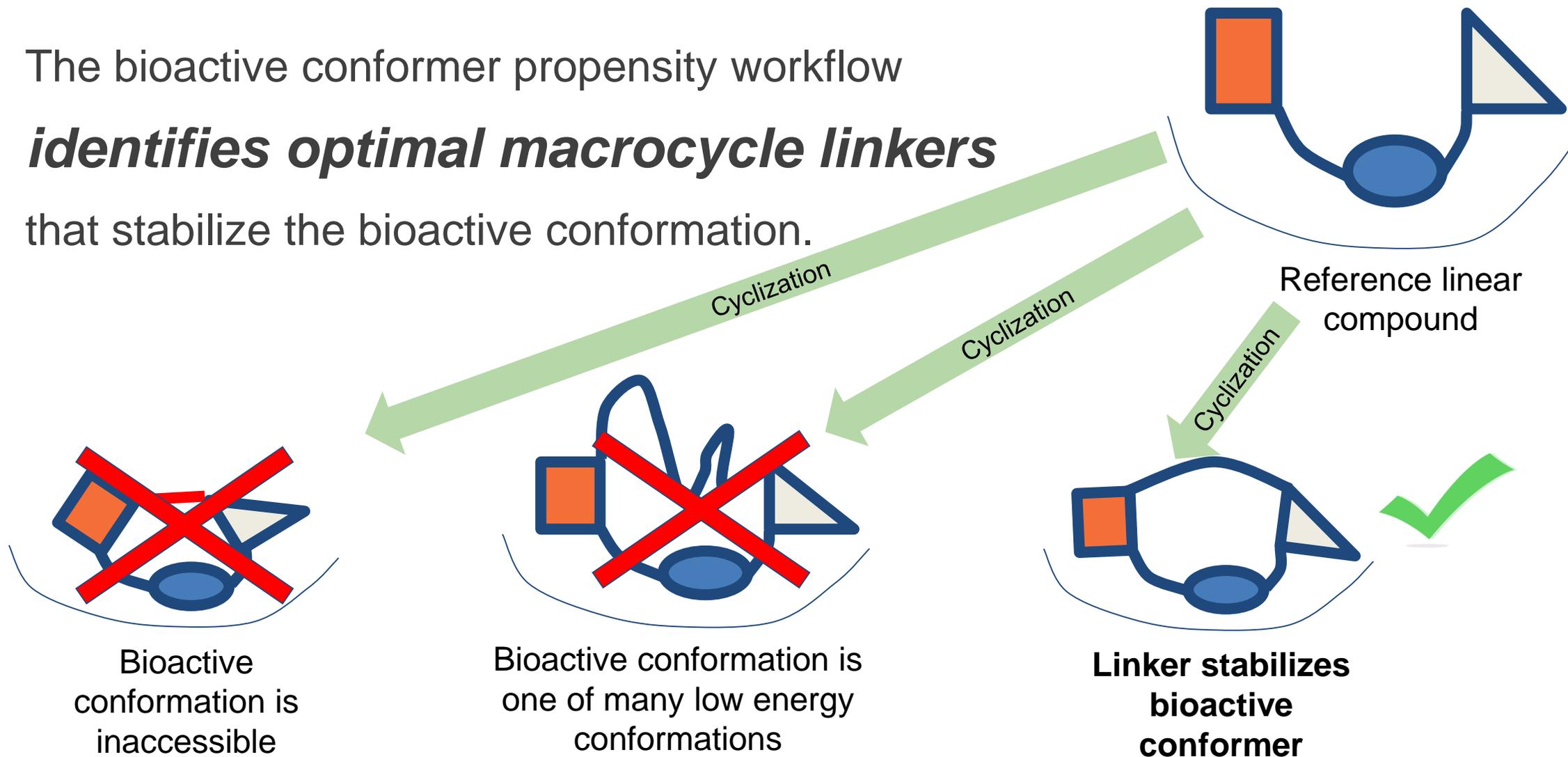
Predicting Stable Cyclizations of Linear Ligands

Cyclization tool: Macrocycle Bioactive Conformer Propensity

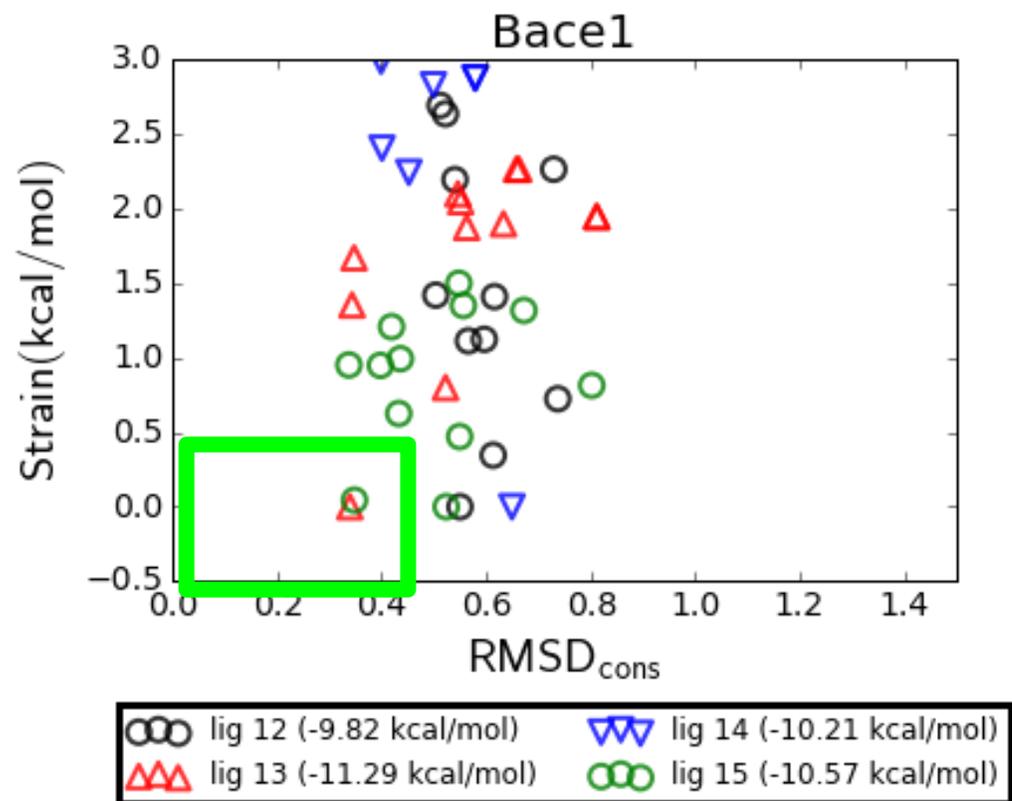
The bioactive conformer propensity workflow

identifies optimal macrocycle linkers

that stabilize the bioactive conformation.



Strain vs RMSD of “Conserved” Region for Sampled Conformers

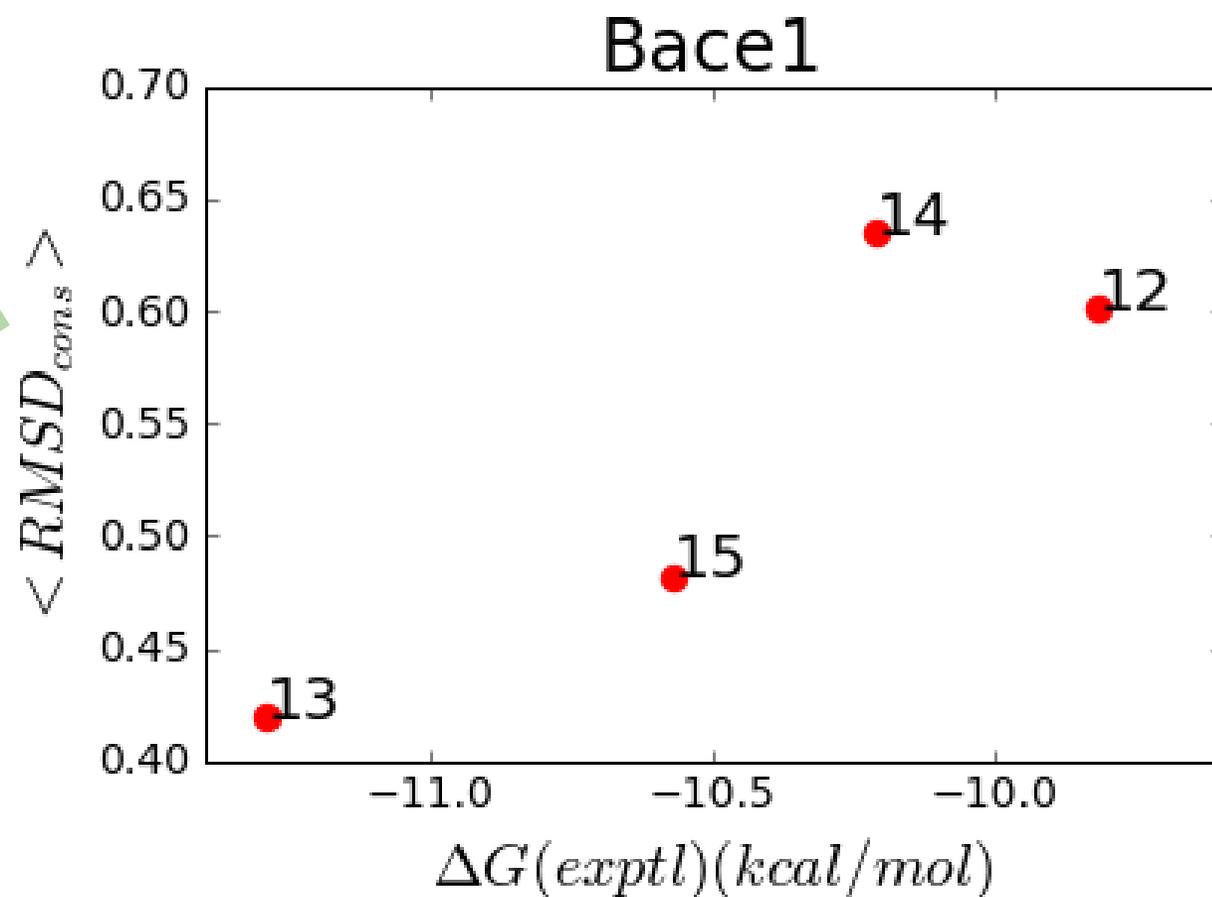
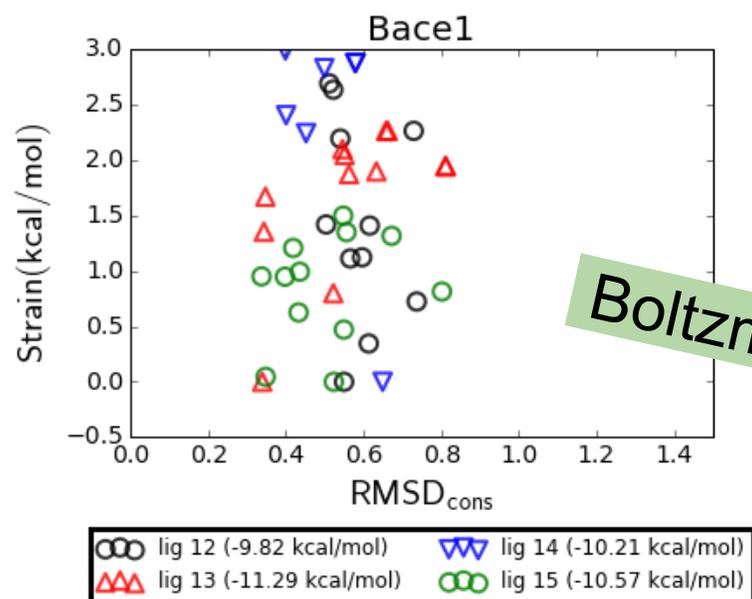


1. **Sample each ligand using PrimeMCS**
2. **Calculate the RMSDs** of the substructure important for activity for each member of the ensemble to the interacting portion of the known active ligand. (RMSD and relative energy are plotted).
3. **Calculate the expected RMSD** by Boltzmann weighting over the ensemble (next slide) to quantify *propensity for the bioactive conformer*

Prime energy vs RMSD of Bioactive SMARTS

Examined Case: Huang Y, Strobel ED, Ho CY et al. Bioorg. Med. Chem. Lett. 20(10), 3158–3160 (2010).

$\langle RMSD_{cons} \rangle$ Simplifies Conformational Propensity



Expected RMSD (right) of bioactive region selects* more active macrocycles due to their more stable bioactive conformations.

*Similar performance for additional systems in

A 3D molecular simulation showing a protein structure represented by grey ribbons. A large, orange, textured macrocyclic molecule is bound to a specific pocket of the protein. The text "Predicting Macrocycle Binding Modes" is overlaid in white on the protein structure.

Predicting Macrocycle Binding Modes

Integrating Prime Macrocycle Sampling with Glide

PrimeMCS

Efficiently samples macrocycle ring conformations

Doesn't directly account for environmental effects

Glide docking

Efficiently accounts for receptor environment

Relies on templates for ring conformations

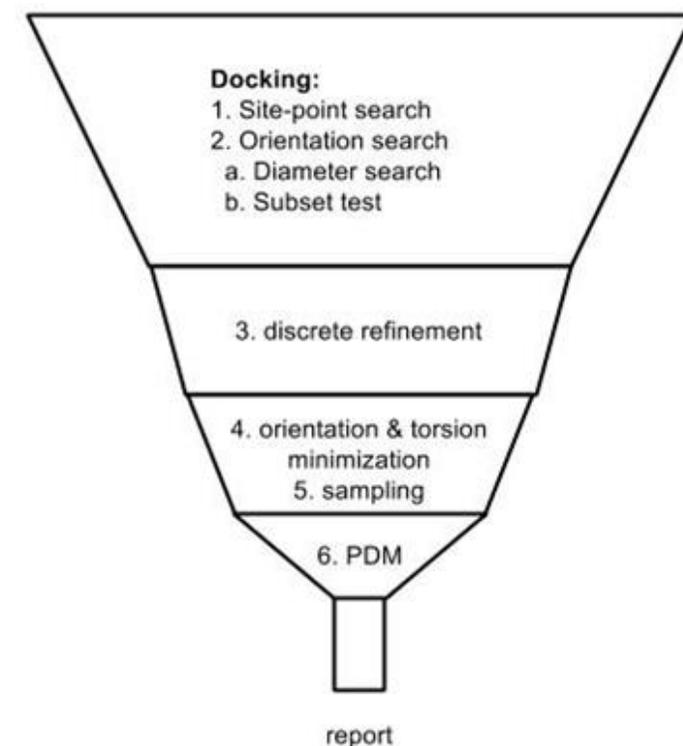
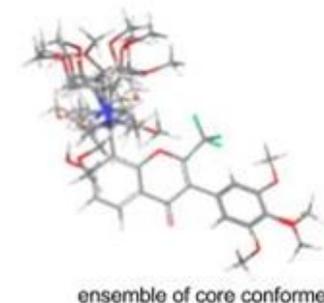
Glide “macrocycle mode”

Generates ring templates using PrimeMCS “on-the-fly” for contextual sampling

Sampling and docking parameters optimized for macrocycle docking

Integrating PrimeMCS into Glide

- Glide uses a filtering-based workflow to go from conformers to poses
- For macrocycle **PrimeMCS** is used to generate on the fly ring templates to be utilized in the initial core conformer ensemble generation (confgen)
- We use an “**expanded funnel**” to reflect the additional conformational and pose complexity added by macrocycles

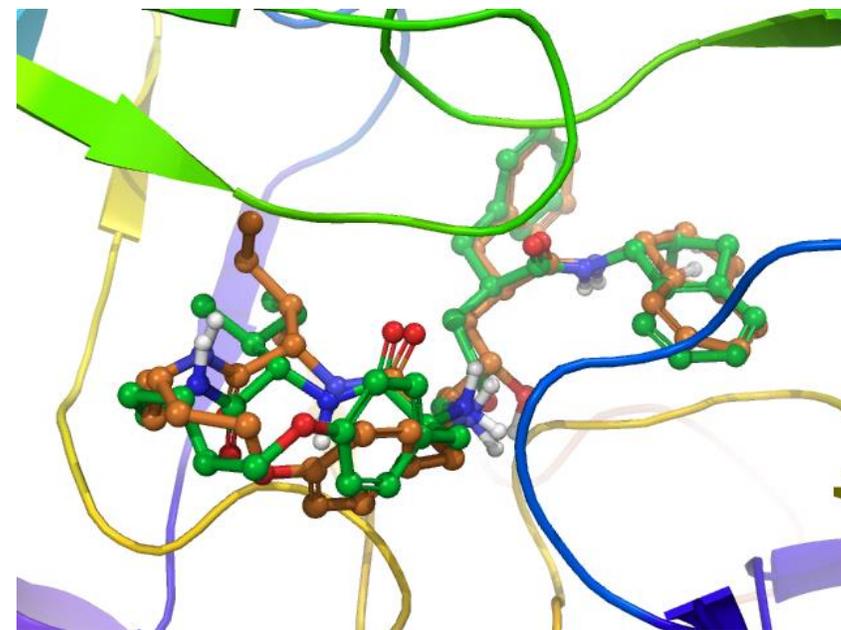


PrimeMCS-integrated Glide Docking Results

Dataset of 67 cocrystallized macrocycles, median 16 backbone atoms

PrimeMCS-based Glide self-docking with no restraints **found top poses under 2.0 Å 70% of the time**, significantly better than with rigid rings, but not as good as docking the native conformation.

	Native conformation	Rigid ring	PrimeMCS-based
% Top pose under 2.0Å	91%	43%	70%
Median Serial CPU time	~1m	~1m	30m



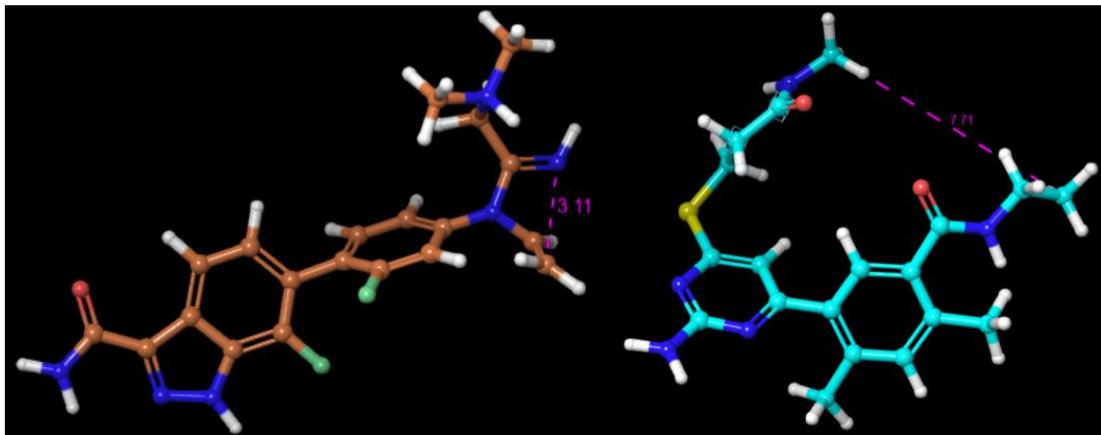
1d4k 1.1Å docked structure

A 3D molecular model showing a protein structure represented by grey ribbons. A large, orange, semi-transparent surface represents a binding pocket or macrocycle. Inside this pocket, a ligand molecule is shown in a stick representation with blue, grey, and yellow atoms. The text "Macrocycle FEP+" is overlaid in white on the orange surface.

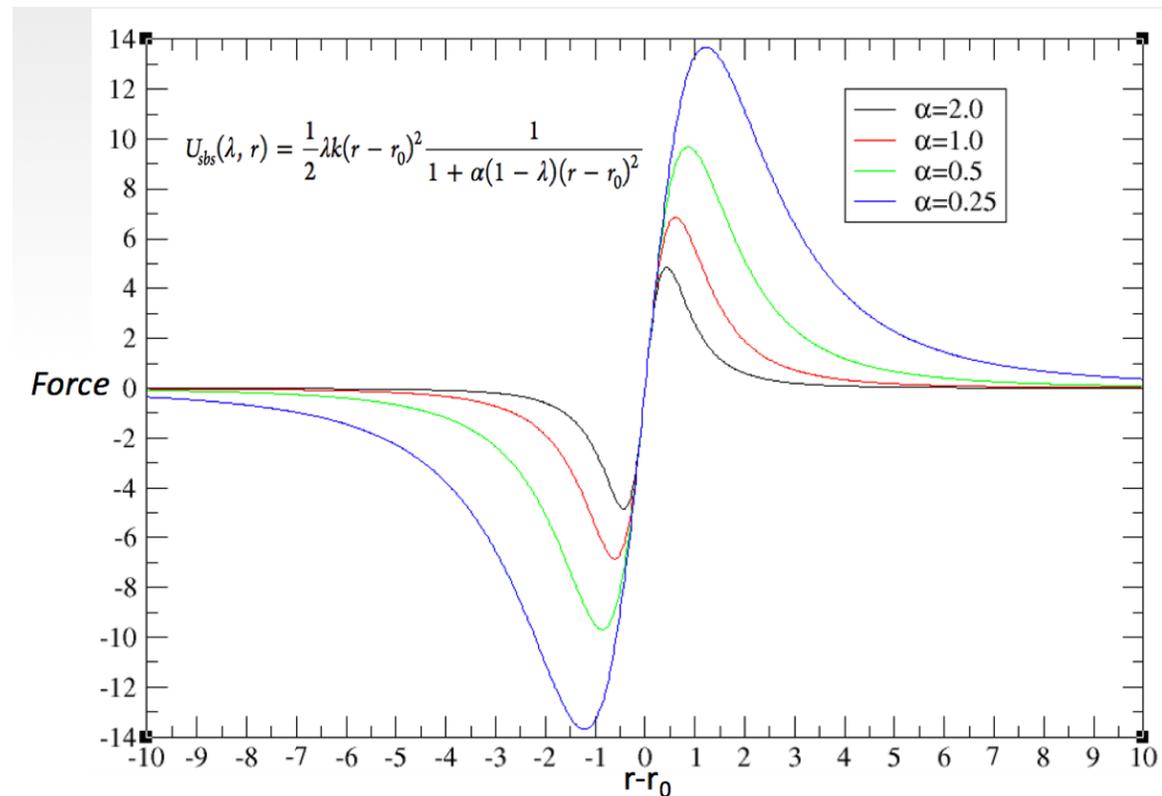
Macrocycle FEP+

Macrocycle FEP+

- Core-hopping technology (soft bond scaling) has enabled macrocyclization reactions in FEP+
- Macrocycles are automatically detected and run with optimal scaling parameters



Bonds can be formed even across long distances

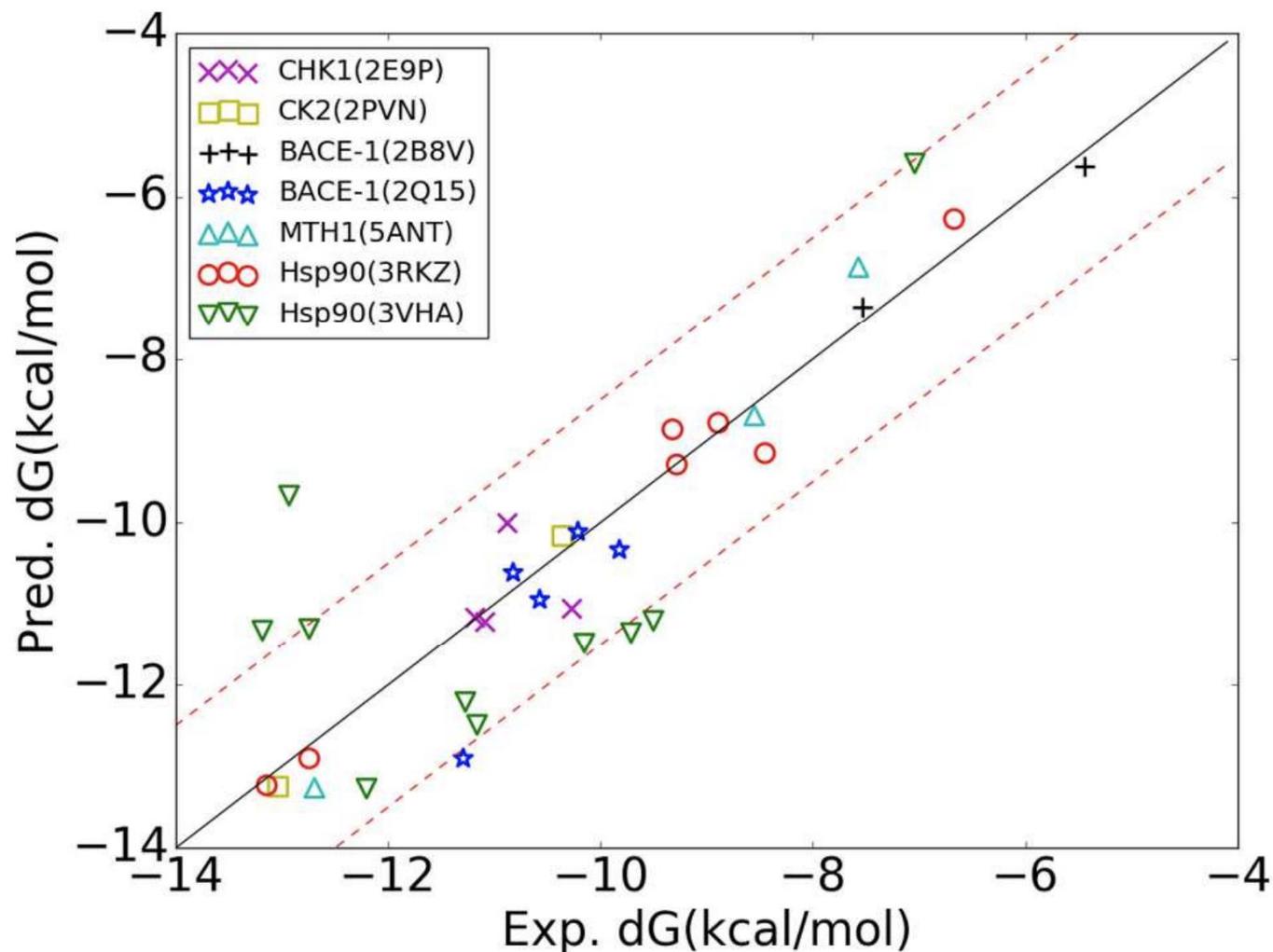


Corresponding force of scaled soft bond potential (above) enables smooth macrocycle bond formation across FEP+ lambda schedule

Macrocycle FEP+ Retrospective Study Results

- Seven retrospective cases of macrocyclization
- $\Delta\Delta G$
 - MUE: 0.71
 - RMSE 0.92

Macrocycle FEP+ currently achieves accuracy on par with small molecule FEP+

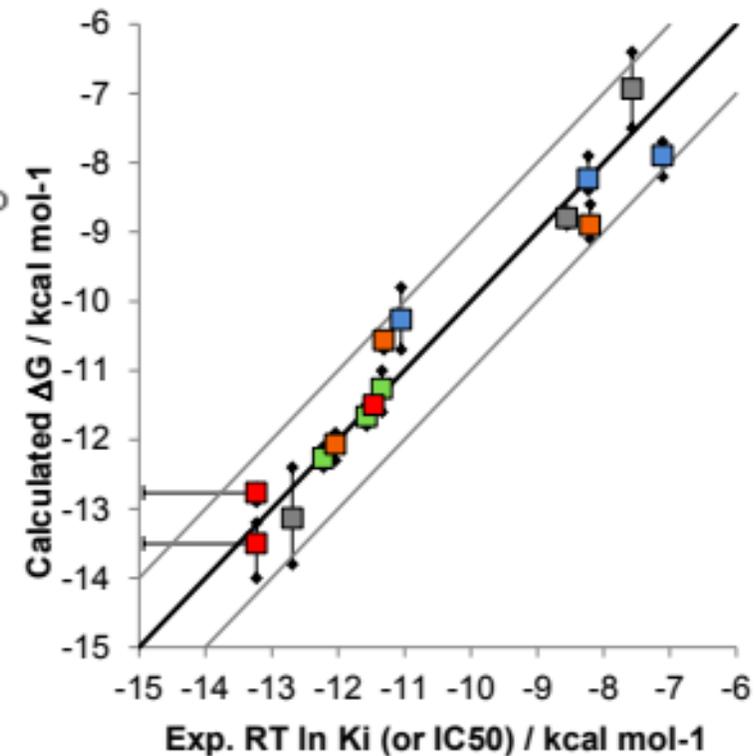
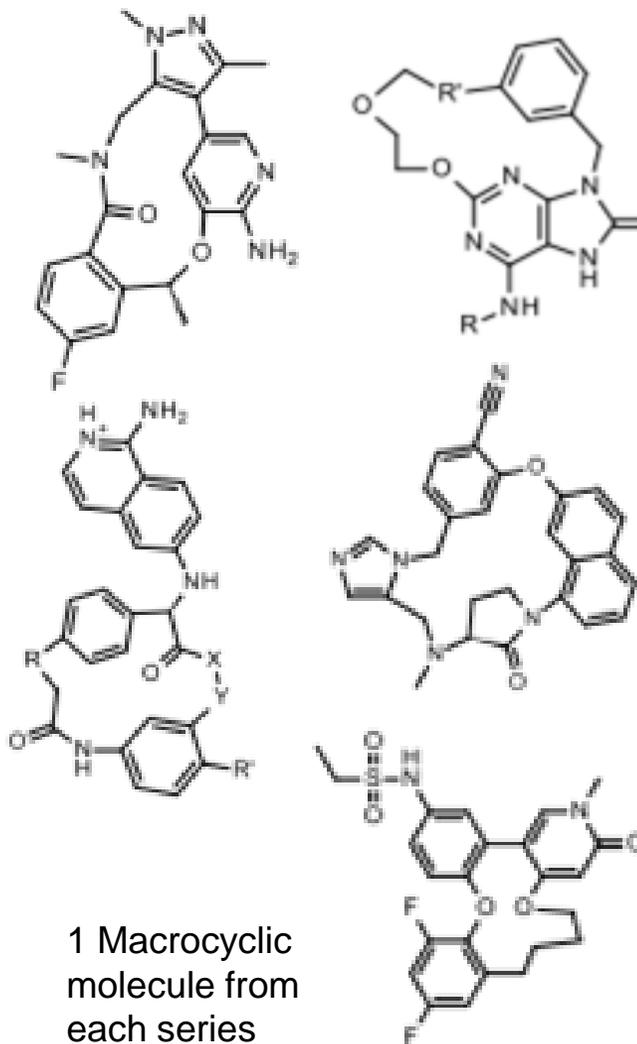


Macrocycle FEP+ at Bayer

The effect of macrocyclization on binding affinity was predicted for 5 diverse pharmaceutically-relevant targets.

FEP+ was able to predict all binding affinities to within 1kcal/mol

Many series involved a mix of macrocyclic and linear molecules



Calculated versus experimental binding affinities for all targets. Mean (squares), minimum and maximum values. Colored by molecules in the same series.

A molecular structure visualization featuring a protein backbone represented by grey ribbons. A large, orange, semi-transparent surface model of a ligand is bound to the protein. A stick model of the ligand is overlaid on the surface, showing a complex ring system with blue and yellow atoms. The background is dark grey.

Macrocycle Tools Summary

Summary of Schrödinger's Macrocycle Design Tools

- Schrödinger tools offer comprehensive capabilities for macrocycles
- Prime-MCSs sampling enables many workflows
 - Macrocycle bioactive conformer stability calculations
 - Docking within Glide
 - Membrane permeability predictions
- FEP+ for macrocycles
- We are actively developing macrocycle tools and workflows!

Acknowledgements

- Sampling

- BMS
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- FEP+

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- Robert Abel
- Yuqing Deng

- Docking

- Ivan Tubert-Brohman

- Prime

- Ed Miller

- Infrastructure

- Tor Colvin
- Dan Nealschneider



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